

“The Problem of Spatial Autocorrelation:” forty years on

Roger Bivand

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1 Introduction

Cliff and Ord (1969), published forty years ago, marked a turning point in the treatment of spatial autocorrelation in quantitative geography. It provided the framework needed by any applied researcher to attempt an implementation for a different system, possibly using a different programming language. In this spirit, here we examine how spatial weights have been represented in implementations and may be reproduced, how the tabulated results in the paper may be reproduced, and how they may be extended to cover simulation.

One of the major assertions of Cliff and Ord (1969) is that their statistic advances the measurement of spatial autocorrelation with respect to Moran (1950) and Geary (1954) because a more general specification of spatial weights could be used. This more general form has implications both for the preparation of the weights themselves, and for the calculation of the measures. We will look at spatial weights first, before moving on to consider the measures presented in the paper and some of their subsequent developments. Before doing this, we will put together a data set matching that used in Cliff and Ord (1969). They provide tabulated data for the counties of the Irish Republic, but omit Dublin from analyses. A shapefile included in this package, kindly made available by Michael Tiefelsdorf, is used as a starting point:

```
> library(spdep)
> fn <- system.file("etc/shapes/eire.shp", package = "spdep")[1]
> prj <- CRS("+proj=utm +zone=30 +units=km")
> eire <- readShapeSpatial(fn, ID = "names", proj4string = prj)
> class(eire)
[1] "SpatialPolygonsDataFrame"
attr(,"package")
[1] "sp"
> names(eire)
[1] "A"          "towns"      "pale"       "size"       "ROADACC"    "OWNCONS"    "POPCHG"    "RETSALE"
[9] "INCOME"     "names"
```

and read into a `SpatialPolygonsDataFrame` — classes used for handling spatial data in R are fully described in Bivand, Pebesma, and Gómez-Rubio (2008). To this we need to add the data tabulated in the paper in Table 2,¹ p. 40, here in the form of a text file with added rainfall values from Table 9, p. 49:

```
> fn <- system.file("etc/misc/geary_eire.txt", package = "spdep")[1]
> ge <- read.table(fn, header = TRUE)
> names(ge)
```

¹cropped scans of tables are available from <http://spatial.nhh.no/R/etc/C069-PNGs.zip>.

```
[1] "serlet"      "county"      "pagval2_10"   "pagval10_50"  "pagval50p"
[6] "cowspacre"   "ocattlepacre" "pigspacre"    "sheeppacre"   "townvillp"
[11] "carspcap"    "radiopcap"   "retailpcap"   "psinglem30_34" "rainfall"
```

Since we assigned the county names as feature identifiers when reading the shapefiles, we do the same with the extra data, and combine the objects:

```
> row.names(ge) <- as.character(ge$county)
> all.equal(row.names(ge), row.names(eire))
[1] TRUE
> eire_ge <- spCbind(eire, ge)
```

Finally, we need to drop the Dublin county omitted in the analyses conducted in Cliff and Ord (1969):

```
> eire_ge1 <- eire_ge[!(row.names(eire_ge) %in% "Dublin"), ]
> length(row.names(eire_ge1))
[1] 25
```

To double-check our data, let us calculate the sample Beta coefficients, using the formulae given in the paper for sample moments:

```
> skewness <- function(z) {
+   z <- scale(z, scale = FALSE)
+   ((sum(z^3)/length(z))^2)/((sum(z^2)/length(z))^3)
+ }
> kurtosis <- function(z) {
+   z <- scale(z, scale = FALSE)
+   (sum(z^4)/length(z))/((sum(z^2)/length(z))^2)
+ }
```

These differ somewhat from the ways in which skewness and kurtosis are computed in modern statistical software, see for example Joanes and Gill (1998). However, for our purposes, they let us reproduce Table 3, p. 42:

```
> print(sapply(as(eire_ge1, "data.frame")[13:24], skewness), digits = 3)

pagval2_10 pagval10_50 pagval50p cowspacre ocattlepacre pigspacre
1.675429 1.294978 0.000382 1.682094 0.086267 1.138387
sheeppacre townvillp carspcap radiopcap retailpcap psinglem30_34
1.842362 0.472748 0.011111 0.342805 0.002765 0.068169

> print(sapply(as(eire_ge1, "data.frame")[13:24], kurtosis), digits = 4)

pagval2_10 pagval10_50 pagval50p cowspacre ocattlepacre pigspacre
3.790 4.331 1.508 4.294 2.985 3.754
sheeppacre townvillp carspcap radiopcap retailpcap psinglem30_34
4.527 2.619 1.865 2.730 2.188 2.034

> print(sapply(as(eire_ge1, "data.frame")[c(13, 16, 18, 19)], function(x) skewness(log(x))),
+ digits = 3)

pagval2_10 cowspacre pigspacre sheeppacre
0.68801 0.17875 0.00767 0.04184

> print(sapply(as(eire_ge1, "data.frame")[c(13, 16, 18, 19)], function(x) kurtosis(log(x))),
+ digits = 4)

pagval2_10 cowspacre pigspacre sheeppacre
2.883 2.799 2.212 2.421
```

Using the tabulated value of 23.6 for the percentage of agricultural holdings above 50 in 1950 in Waterford, the skewness and kurtosis cannot be reproduced, but by comparison with the *irishdata* dataset in **ade4**, it turns out that the value should rather be 26.6, which yields the tabulated skewness and kurtosis values.

Before going on, the variables considered are presented in Table 1.

Table 1: Description of variables in the Geary data set.

variable	description
pagval2_10	Percentage number agricultural holdings in valuation group £2–£10 (1950)
pagval10_50	Percentage number agricultural holdings in valuation group £10–£50 (1950)
pagval50p	Percentage number agricultural holdings in valuation group above £50 (1950)
cowspacre	Milch cows per 1000 acres crops and pasture (1952)
ocattlepacre	Other cattle per 1000 acres crops and pasture (1952)
pigspacre	Pigs per 1000 acres crops and pasture (1952)
sheeppacre	Sheep per 1000 acres crops and pasture (1952)
townvillp	Town and village population as percentage of total (1951)
carspcap	Private cars registered per 1000 population (1952)
radiopcap	Radio licences per 1000 population (1952)
retailpcap	Retail sales £ per person (1951)
psinglem30_34	Single males as percentage of all males aged 30–34 (1951)
rainfall	Average of rainfall for stations in Ireland, 1916–1950, mm

2 Spatial weights

As a basis for comparison, we will first read the unstandardised weighting matrix given in Table A1, p. 54, of the paper, reading a file corrected for the misprint giving O rather than D as a neighbour of V:

```
> fn <- system.file("etc/misc/unstand_sn.txt", package = "spdep")[1]
> unstand <- read.table(fn, header = TRUE)
> summary(unstand)
```

```
      from      to      weight
V      : 8  V      : 8  Min.    :0.000600
S      : 7  S      : 7  1st Qu.:0.003225
T      : 7  T      : 7  Median :0.007550
Q      : 6  Q      : 6  Mean    :0.007705
A      : 5  A      : 5  3rd Qu.:0.010225
B      : 5  B      : 5  Max.    :0.032400
(Other):72  (Other):72
```

In the file, the counties are represented by their serial letters, so ordering and conversion to integer index representation is required to reach a representation similar to that of the S-PLUS SpatialStats module for spatial neighbours:

```
> class(unstand) <- c("spatial.neighbour", class(unstand))
> of <- ordered(unstand$from)
> attr(unstand, "region.id") <- levels(of)
> unstand$from <- as.integer(of)
> unstand$to <- as.integer(ordered(unstand$to))
> attr(unstand, "n") <- length(unique(unstand$from))
```

Having done this, we can change its representation to a `listw` object, assigning an appropriate style (generalised binary) for unstandardised values:

```
> lw_unstand <- sn2listw(unstand)
> lw_unstand$style <- "B"
> lw_unstand
```

Characteristics of weights list object:

Neighbour list object:

Number of regions: 25

Number of nonzero links: 110

Percentage nonzero weights: 17.6

Average number of links: 4.4

Weights style: B

Weights constants summary:

```

      n  nn    S0      S1      S2
B 25 625 0.8476 0.01871808 0.1229232

```

Note that the values of S0, S1, and S2 correspond closely with those given on page 42 of the paper, 0.84688672, 0.01869986 and 0.12267319. The discrepancies appear to be due to rounding in the printed table of weights.

The contiguous neighbours represented in this object ought to match those found using `poly2nb`. However, we see that the reproduced contiguities have a smaller link count:

```

> nb <- poly2nb(eire_ge1)
> nb

Neighbour list object:
Number of regions: 25
Number of nonzero links: 108
Percentage nonzero weights: 17.28
Average number of links: 4.32

```

The missing link is between Clare and Kerry, perhaps by the Tarbert–Killimer ferry, as Figure 1 shows:

```

> xx <- diffnb(nb, lw_unstand$neighbours, verbose = TRUE)

Neighbour difference for region id: Clare in relation to id: Kerry
Neighbour difference for region id: Kerry in relation to id: Clare

> plot(eire_ge1, border = "grey60")
> plot(nb, coordinates(eire_ge1), add = TRUE, pch = ".", lwd = 2)
> plot(xx, coordinates(eire_ge1), add = TRUE, pch = ".", lwd = 2, col = 3)

```

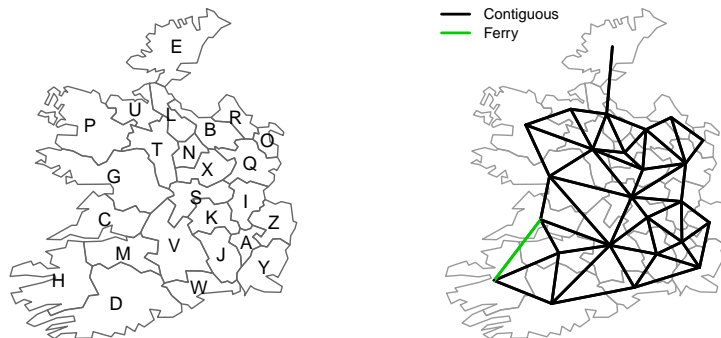


Figure 1: County boundaries and contiguities

An attempt has also been made to reproduce the generalised weights for 25 Irish counties reported by Cliff and Ord (1969), after Dublin is omitted. Reproducing the inverse distance component d_{ij}^{-1} of the generalised weights $d_{ij}^{-1}\beta_{i(j)}$ is eased by the statement in Cliff and Ord (1973, p. 55) that the points chosen to represent the counties were their “geographic centres,” so not very different from the centroids yielded by applying a chosen computational geometry function. The distance metric is not given, and may have been in kilometers or miles — both were tried, but the results were not sensitive to the difference as it applies equally across the weights; miles are used here.

Computing the proportion of shared distance measure $\beta_{i(j)}$ is harder, because it requires the availability of the full topology of the input polygons. Bivand, Pebesma, and Gómez-Rubio (2008, p. 244) show how to employ the `vect2neigh` function (written by Markus Neteler) in the R **spgrass6** package when using GRASS GIS vector handling to create a full topology from spaghetti vector data and to extract border segment lengths; a similar approach also is mentioned there using ArcGIS coverages for the same purpose. GRASS was used to create the topology, and next the proportion of shared distance measure was calculated.

```
> library(maptools)
> SG <- Sobj_SpatialGrid(eire_ge1)$SG
> library(spgrass6)
> grass_home <- "/home/rsb/topics/grass/g64/grass-6.4.0svn"
> initGRASS(grass_home, home = tempdir(), SG = SG, override = TRUE)
> writeVECT6(eire_ge1, "eire", v.in.ogr_flags = c("o", "overwrite"))
> res <- vect2neigh("eire", ID = "serlet")

> grass_borders <- sn2listw(res)
> raw_borders <- grass_borders$weights
> int_tot <- attr(res, "total") - attr(res, "external")
> prop_borders <- lapply(1:length(int_tot), function(i) raw_borders[[i]]/int_tot[i])
> dlist <- nbdistts(grass_borders$neighbours, coordinates(eire_ge1))
> inv_dlist <- lapply(dlist, function(x) 1/(x/1.609344))
> combo_km <- lapply(1:length(inv_dlist), function(i) inv_dlist[[i]] *
+   prop_borders[[i]])
> combo_km_lw <- nb2listw(grass_borders$neighbours, glist = combo_km,
+   style = "B")
> summary(combo_km_lw)

Characteristics of weights list object:
Neighbour list object:
Number of regions: 25
Number of nonzero links: 108
Percentage nonzero weights: 17.28
Average number of links: 4.32
Link number distribution:

1 2 3 4 5 6 7 8
1 2 5 5 8 1 2 1
1 least connected region:
E with 1 link
1 most connected region:
V with 8 links

Weights style: B
Weights constants summary:
  n  nn    S0    S1.5    S2
B 25 625 0.9083141 0.02191843 0.1427745
```

To compare, we need to remove the Tarbert–Killimer ferry link manually, and view the summary of the original weights, as well as a correlation coefficient between these and the reconstructed weights. Naturally, unless the boundary coordinates used here are identical with those in the original analysis, presumably measured by hand, small differences will occur.

```
> red_lw_unstand <- lw_unstand
> Clare <- which(attr(lw_unstand, "region.id") == "C")
> Kerry <- which(attr(lw_unstand, "region.id") == "H")
> Kerry_in_Clare <- which(lw_unstand$neighbours[[Clare]] == Kerry)
> Clare_in_Kerry <- which(lw_unstand$neighbours[[Kerry]] == Clare)
> red_lw_unstand$neighbours[[Clare]] <- red_lw_unstand$neighbours[[Clare]][-Kerry_in_Clare]
> red_lw_unstand$neighbours[[Kerry]] <- red_lw_unstand$neighbours[[Kerry]][-Clare_in_Kerry]
```

```

> red_lw_unstand$weights[[Clare]] <- red_lw_unstand$weights[[Clare]][-Kerry_in_Clar]
> red_lw_unstand$weights[[Kerry]] <- red_lw_unstand$weights[[Kerry]][-Clare_in_Kerry]
> summary(red_lw_unstand)

Characteristics of weights list object:
Neighbour list object:
Number of regions: 25
Number of nonzero links: 108
Percentage nonzero weights: 17.28
Average number of links: 4.32
Link number distribution:

1 2 3 4 5 6 7 8
1 2 5 5 8 1 2 1
1 least connected region:
E with 1 link
1 most connected region:
V with 8 links

Weights style: B
Weights constants summary:
      n  nn   S0      S1      S2
B 25 625 0.8437 0.01870287 0.1222501

> cor(unlist(red_lw_unstand$weights), unlist(combo_km_lw$weights))

[1] 0.969543

```

Even though the differences in the general weights, for identical contiguities, are so small, the consequences for the measure of spatial autocorrelation are substantial. Here we use the fifth variable, other cattle per 1000 acres crops and pasture (1952), and see that the reconstructed weights seem to “reveal” more autocorrelation than the original weights.

```

> flatten <- function(x, digits = 3, statistic = "I") {
+   res <- c(format(x$estimate, digits = digits), format(x$statistic,
+     digits = digits), format.pval(x$p.value, digits = digits))
+   res <- matrix(res, ncol = length(res))
+   colnames(res) <- paste(c("", "E", "V", "SD_", "P_"), "I", sep = "")
+   rownames(res) <- deparse(substitute(x))
+   res
+ }

> `reconstructed weights` <- moran.test(eire_ge1$cattlepacre, combo_km_lw)
> `original weights` <- moran.test(eire_ge1$cattlepacre, red_lw_unstand)
> print(rbind(flatten(`reconstructed weights`), flatten(`original weights`)),
+   quote = FALSE)

```

	I	EI	VI	SD_I	P_I
reconstructed weights	0.3203	-0.0417	0.0225	2.41	0.00792
original weights	0.2779	-0.0417	0.0223	2.14	0.0161

3 Measures of spatial autocorrelation

Our targets for reproduction are Tables 4 and 5 in Cliff and Ord (1969, pp. 43–44), the first containing standard deviates under normality and randomisation for the original Moran measure with binary weights, the original Geary measure with binary weights, the proposed measure with unstandardised general weights, and the proposed measure with row-standardised general weights. In addition, four variables were log-transformed on the basis of the skewness and kurtosis results presented above. We carry out the transformation of these variables, and generate additional binary and row-standardised general spatial weights objects — note that the weights constants for the

row-standardised general weights agree with those given on p. 42 in the paper, after allowing for small differences due to rounding in the weights values displayed in the paper (p. 54):

```
> eire_ge1$ln_pagval2_10 <- log(eire_ge1$pagval2_10)
> eire_ge1$ln_cowspacre <- log(eire_ge1$cowspacre)
> eire_ge1$ln_pigspacre <- log(eire_ge1$pigspacre)
> eire_ge1$ln_sheepacre <- log(eire_ge1$sheepacre)
> vars <- c("pagval2_10", "ln_pagval2_10", "pagval10_50", "pagval50p",
+ "cowspacre", "ln_cowspacre", "ocattlepacre", "pigspacre", "ln_pigspacre",
+ "sheepacre", "ln_sheepacre", "townvillp", "carspcap", "radiopcap",
+ "retailpcap", "psinglem30_34")
> nb_B <- nb2listw(lw_unstand$neighbours, style = "B")
> nb_B
```

```
Characteristics of weights list object:
Neighbour list object:
Number of regions: 25
Number of nonzero links: 110
Percentage nonzero weights: 17.6
Average number of links: 4.4
```

```
Weights style: B
Weights constants summary:
  n nn S0 S1 S2
B 25 625 110 220 2176
```

```
> lw_std <- nb2listw(lw_unstand$neighbours, glist = lw_unstand$weights,
+ style = "W")
> lw_std
```

```
Characteristics of weights list object:
Neighbour list object:
Number of regions: 25
Number of nonzero links: 110
Percentage nonzero weights: 17.6
Average number of links: 4.4
```

```
Weights style: W
Weights constants summary:
  n nn S0 S1 S2
W 25 625 25 15.84089 103.6197
```

The standard representation of the measures is:

$$I = \frac{n}{\sum_{i=1}^n \sum_{j=1}^n w_{ij}} \frac{\sum_{i=1}^n \sum_{j=1}^n w_{ij} (x_i - \bar{x})(x_j - \bar{x})}{\sum_{i=1}^n (x_i - \bar{x})^2}$$

for Moran's I — in the paper termed the proposed statistic, and for Geary's C :

$$C = \frac{(n-1)}{2 \sum_{i=1}^n \sum_{j=1}^n w_{ij}} \frac{\sum_{i=1}^n \sum_{j=1}^n w_{ij} (x_i - x_j)^2}{\sum_{i=1}^n (x_i - \bar{x})^2}$$

where $x_i, i = 1, \dots, n$ are n observations on the numeric variable of interest, and w_{ij} are the spatial weights. In order to reproduce the standard deviates given in the paper, it is sufficient to apply `moran.test` to the variables with three different spatial weights objects, and two different values of the `randomisation=` argument. In addition, `geary.test` is applied to a single spatial weights objects, and two different values of the `randomisation=` argument.

```

> system.time({
+   MoranN <- lapply(vars, function(x) moran.test(eire_ge1[[x]], listw = nb_B,
+     randomisation = FALSE))
+   MoranR <- lapply(vars, function(x) moran.test(eire_ge1[[x]], listw = nb_B,
+     randomisation = TRUE))
+   GearyN <- lapply(vars, function(x) geary.test(eire_ge1[[x]], listw = nb_B,
+     randomisation = FALSE))
+   GearyR <- lapply(vars, function(x) geary.test(eire_ge1[[x]], listw = nb_B,
+     randomisation = TRUE))
+   Prop_unstdN <- lapply(vars, function(x) moran.test(eire_ge1[[x]],
+     listw = lw_unstand, randomisation = FALSE))
+   Prop_unstdR <- lapply(vars, function(x) moran.test(eire_ge1[[x]],
+     listw = lw_unstand, randomisation = TRUE))
+   Prop_stdN <- lapply(vars, function(x) moran.test(eire_ge1[[x]],
+     listw = lw_std, randomisation = FALSE))
+   Prop_stdR <- lapply(vars, function(x) moran.test(eire_ge1[[x]],
+     listw = lw_std, randomisation = TRUE))
+ })

      user  system elapsed
0.906   0.004   0.933

> res <- sapply(c("MoranN", "MoranR", "GearyN", "GearyR", "Prop_unstdN",
+   "Prop_unstdR", "Prop_stdN", "Prop_stdR"), function(x) sapply(get(x),
+   "[", "statistic"))
> rownames(res) <- vars
> ores <- res[, c(1, 2, 5:8)]

```

In order to conduct 8 different tests on 16 variables, we use `lapply` on the list of variables in the specified order, then `sapply` on a list of output objects by name to generate a table in the same row and column order as the original (we save a copy of six columns for comparison with bootstrap results below):

```

> print(formatC(res, format = "f", digits = 4), quote = FALSE)

```

	MoranN	MoranR	GearyN	GearyR	Prop_unstdN	Prop_unstdR	Prop_stdN	Prop_stdR
pagval2_10	3.7851	3.8779	4.3142	3.9016	3.3307	3.4159	3.9276	4.0292
ln_pagval2_10	4.0965	4.1074	4.0841	4.0343	3.5795	3.5894	4.1278	4.1393
pagval10_50	1.0899	1.1316	2.7511	2.3760	1.3348	1.3882	1.5127	1.5738
pagval50p	5.2011	5.0555	4.0178	4.7194	4.6604	4.5247	4.8823	4.7387
cowspacre	5.1969	5.3907	4.3531	3.7709	4.1379	4.2991	4.7274	4.9135
ln_cowspacre	5.2420	5.2455	3.9211	3.9085	4.2007	4.2037	4.6532	4.6566
ocattlepacre	0.5565	0.5593	-0.1707	-0.1668	2.1366	2.1478	1.9219	1.9320
pigspacre	2.4807	2.5393	2.2928	2.0802	2.8312	2.9010	3.1908	3.2703
ln_pigspacre	2.3015	2.2724	2.0520	2.1893	2.5171	2.4839	2.8460	2.8081
sheeppacre	1.0188	1.0630	0.8689	0.7387	1.7398	1.8187	1.4792	1.5470
ln_sheeppacre	1.2930	1.2827	1.5156	1.5767	2.3708	2.3511	2.0374	2.0203
townvillp	2.2759	2.2681	2.5475	2.5902	1.2148	1.2104	1.6275	1.6216
carspcap	4.4927	4.4015	3.2247	3.5992	3.8897	3.8075	4.1826	4.0934
radiopcap	0.3156	0.3153	1.2294	1.2348	0.5915	0.5909	0.7857	0.7849
retailpcap	3.4985	3.4524	3.1303	3.3497	2.9291	2.8888	3.0346	2.9926
psinglem30_34	2.7349	2.6895	2.3382	2.5519	2.7541	2.7065	2.7078	2.6605

The values of the standard deviates agree with those in Table 4 in the original paper, with the exception of those for the proposed statistic with standardised weights under normality for all untransformed variables. We can see what has happened by substituting the weights constants for the standardised weights with those for unstandardised weights:

```

> wc_unstd <- spweights.constants(lw_unstand)
> wrong_N_sqVI <- sqrt((wc_unstd$nn * wc_unstd$S1 - wc_unstd$N * wc_unstd$S2 +
+   3 * wc_unstd$S0 * wc_unstd$S0)/((wc_unstd$nn - 1) * wc_unstd$S0 *
+   wc_unstd$S0) - ((-1/(wc_unstd$N - 1))^2))
> raw_data <- grep("~ln_", vars, invert = TRUE)

```



```

> I <- sapply(Prop_stdN, function(x) x$estimate[1])[raw_data]
> EI <- sapply(Prop_stdN, function(x) x$estimate[2])[raw_data]
> res <- (I - EI)/wrong_N_sqVI
> names(res) <- vars[raw_data]
> print(formatC(res, format = "f", digits = 4), quote = FALSE)

```

	pagval2_10	pagval10_50	pagval50p	cowspacre	ocattlepacre	pigspacre
	3.8836	1.4957	4.8276	4.6744	1.9003	3.1550
	sheeppacre	townvillp	carspcap	radiopcap	retailpcap	psinglem30_34
	1.4627	1.6093	4.1357	0.7769	3.0006	2.6774

Next, let us look at Table 5 in the original paper. Here we only tabulate the values of the measures themselves, and, since the expectation is constant for each measure, the square root of the variance of the measure under randomisation — extracting values calculated above:

```

> res <- lapply(c("MoranR", "GearyR", "Prop_unstdR", "Prop_stdR"), function(x) sapply(get(x),
+   function(y) c(y$estimate[1], sqrt(y$estimate[3]))))
> res <- t(do.call("rbind", res))
> colnames(res) <- c("I", "sigma_I", "C", "sigma_C", "unstd_r", "sigma_r",
+   "std_r", "sigma_r")
> rownames(res) <- vars
> print(formatC(res, format = "f", digits = 4), quote = FALSE)

```

	I	sigma_I	C	sigma_C	unstd_r	sigma_r	std_r	sigma_r
pagval2_10	0.4074	0.1158	0.3477	0.1672	0.4559	0.1456	0.5384	0.1440
ln_pagval2_10	0.4444	0.1183	0.3825	0.1531	0.4930	0.1490	0.5680	0.1473
pagval10_50	0.0877	0.1143	0.5840	0.1751	0.1577	0.1436	0.1818	0.1420
pagval50p	0.5754	0.1221	0.3925	0.1287	0.6545	0.1539	0.6795	0.1522
cowspacre	0.5749	0.1144	0.3418	0.1745	0.5764	0.1438	0.6566	0.1421
ln_cowspacre	0.5803	0.1186	0.4071	0.1517	0.5858	0.1493	0.6456	0.1476
ocattlepacre	0.0244	0.1181	1.0258	0.1547	0.2775	0.1486	0.2422	0.1469
pigspacre	0.2527	0.1159	0.6533	0.1667	0.3812	0.1458	0.4296	0.1441
ln_pigspacre	0.2314	0.1202	0.6897	0.1417	0.3343	0.1514	0.3787	0.1497
sheeppacre	0.0792	0.1137	0.8686	0.1778	0.2182	0.1429	0.1768	0.1412
ln_sheeppacre	0.1117	0.1196	0.7708	0.1453	0.3125	0.1506	0.2593	0.1489
townvillp	0.2284	0.1191	0.6148	0.1487	0.1398	0.1499	0.1987	0.1482
carspcap	0.4914	0.1211	0.5124	0.1355	0.5394	0.1526	0.5761	0.1509
radiopcap	-0.0042	0.1188	0.8141	0.1505	0.0467	0.1495	0.0744	0.1478
retailpcap	0.3734	0.1202	0.5267	0.1413	0.3959	0.1515	0.4066	0.1498
psinglem30_34	0.2828	0.1207	0.6465	0.1385	0.3697	0.1520	0.3583	0.1503

The values are as follows, and match the original with the exception of those for the initial version of Moran's I in the first two columns. If we write a function implementing equations 3 and 4:

$$I = \frac{\sum_{i=1}^n \sum_{j=i+1}^n w_{ij} (x_i - \bar{x})(x_j - \bar{x})}{\sum_{i=1}^n (x_i - \bar{x})^2}$$

where crucially the inner summation is over $i + 1 \dots n$, not $1 \dots n$, we can reproduce the values of the measure shown in the original Table 5:

```

> oMoranf <- function(x, nb) {
+   z <- scale(x, scale = FALSE)
+   n <- length(z)
+   glist <- lapply(1:n, function(i) {
+     ii <- nb[[i]]
+     ifelse(ii > i, 1, 0)
+   })
+   lw <- nb2listw(nb, glist = glist, style = "B")
+   wz <- lag(lw, z)
+   I <- (sum(z * wz)/sum(z * z))
+   I

```

```
+ }
> res <- sapply(vars, function(x) oMoranf(eire_ge1[[x]], nb = lw_unstand$neighbours))
> print(formatC(res, format = "f", digits = 4), quote = FALSE)
```

pagval2_10	ln_pagval2_10	pagval10_50	pagval50p	cowspacre	ln_cowspacre
0.8964	0.9776	0.1928	1.2660	1.2649	1.2766
ocattlepacre	pigspacre	ln_pigspacre	sheeppacre	ln_sheeppacre	townvillp
0.0536	0.5559	0.5091	0.1743	0.2458	0.5024
carspcap	radiopcap	retailpcap	psinglem30_34		
1.0811	-0.0093	0.8215	0.6222		

The variance term given in equation 7 in the original paper is for the case of normality, not randomisation; the reference on p. 28 to equation 38 on p. 26 does not permit the reproduction of the values in the second column of Table 5. The variance equation given as equation 1.35 by Cliff and Ord (1973, p. 9) does not do so either, so for the time being it is not possible to say how the tabulated values were computed. Note that since the standard deviances are reproduced correctly, and can be reproduced from the second column values using the measure and its expectance, it is just a matter of establishing which formula was used, but this has so far not proved possible.

4 Simulating measures of spatial autocorrelation

Cliff and Ord (1969) do not conduct simulation experiments, although their sequels do, notably Cliff and Ord (1973), among many others. Simulation studies are necessarily more demanding computationally, especially if spatially autocorrelated variables are to be created, as in Cliff and Ord (1973, pp. 146–153). In the same book, they also report the use of permutation tests, also known as Monte Carlo or Hope hypothesis testing procedures (Cliff and Ord, 1973, pp. 50–52). These procedures provided a way to examine the distribution of the statistic of interest by exchanging at random the observed values between observations, and then comparing the simulated distribution under the null hypothesis of no spatial patterning with the observed value of the statistic in question.

```
> MoranI.boot <- function(var, i, ...) {
+   var <- var[i]
+   return(moran(x = var, ...)$I)
+ }
> Nsim <- function(d, mle) {
+   n <- length(d)
+   rnorm(n, mle$mean, mle$sd)
+ }
> f_bperm <- function(x, nsim, listw) {
+   boot(x, statistic = MoranI.boot, R = nsim, sim = "permutation",
+       listw = listw, n = length(x), S0 = Szero(listw))
+ }
> f_bpara <- function(x, nsim, listw) {
+   boot(x, statistic = MoranI.boot, R = nsim, sim = "parametric", ran.gen = Nsim,
+       mle = list(mean = mean(x), sd = sd(x)), listw = listw, n = length(x),
+       S0 = Szero(listw))
+ }
> nsim <- 4999
> set.seed(1234)
```

First let us define a function `MoranI.boot` just to return the value of Moran's I for variable `var` and permutation index `i`, and a function `Nsim` to generate random samples from the variable of interest assuming Normality. To make it easier to process the variables in turn, we encapsulate calls to `boot` in wrapper functions `f_bperm` and `f_bpara`.

Running 4999 simulations for each of 16 for three different weights specifications and both parametric and permutation bootstrap takes quite a lot of time.

```
> system.time({
+   MoranNb <- lapply(vars, function(x) f_bpara(x = eire_ge1[[x]], nsim = nsim,
+       listw = nb_B))
+   MoranRb <- lapply(vars, function(x) f_bperm(x = eire_ge1[[x]], nsim = nsim,
+       listw = nb_B))
+   Prop_unstdNb <- lapply(vars, function(x) f_bpara(x = eire_ge1[[x]],
+       nsim = nsim, listw = lw_unstand))
+   Prop_unstdRb <- lapply(vars, function(x) f_bperm(x = eire_ge1[[x]],
+       nsim = nsim, listw = lw_unstand))
+   Prop_stdNb <- lapply(vars, function(x) f_bpara(x = eire_ge1[[x]],
+       nsim = nsim, listw = lw_std))
+   Prop_stdRb <- lapply(vars, function(x) f_bperm(x = eire_ge1[[x]],
+       nsim = nsim, listw = lw_std))
+ })

      user system elapsed
156.161    0.053 156.290

> res <- lapply(c("MoranNb", "MoranRb", "Prop_unstdNb", "Prop_unstdRb",
+   "Prop_stdNb", "Prop_stdRb"), function(x) sapply(get(x), function(y) (y$t0 -
+   mean(y$t))/sd(y$t))))
> res <- t(do.call("rbind", res))
> colnames(res) <- c("MoranNb", "MoranRb", "Prop_unstdNb", "Prop_unstdRb",
+   "Prop_stdNb", "Prop_stdRb")
> rownames(res) <- vars
```

We collate the results to compare with the analytical standard deviates under Normality and randomisation, and see that in fact the differences are not at all large, as expressed by the median absolute difference between the tables. We can also see that inferences based on a one-sided $\alpha = 0.05$ cut-off are the same for the analytical and bootstrap approaches. This indicates that we can, in general, rely on the analytical standard deviates, and that bootstrap methods will not help if assumptions underlying the measures are not met.

```
> print(formatC(res, format = "f", digits = 4), quote = FALSE)
```

	MoranNb	MoranRb	Prop_unstdNb	Prop_unstdRb	Prop_stdNb	Prop_stdRb
pagval2_10	3.7863	3.8492	3.3168	3.4630	3.9634	4.0194
ln_pagval2_10	4.1367	4.0862	3.5868	3.6581	4.1702	4.1103
pagval10_50	1.1046	1.1064	1.3070	1.3692	1.4822	1.5870
pagval50p	5.2336	5.0855	4.6790	4.4855	4.9216	4.6215
cowspacre	5.2719	5.4104	4.1738	4.3003	4.7350	4.8861
ln_cowspacre	5.2422	5.1914	4.1386	4.1536	4.6903	4.6230
ocattlepacre	0.5705	0.5671	2.1297	2.1222	1.9411	1.9267
pigspacre	2.5097	2.5290	2.8613	2.8747	3.2262	3.3209
ln_pigspacre	2.2690	2.2327	2.4767	2.4649	2.8330	2.7646
sheeppacre	1.0252	1.0798	1.7742	1.8546	1.4874	1.5987
ln_sheeppacre	1.2730	1.2635	2.3792	2.3089	2.0554	2.0152
townvillp	2.2430	2.2517	1.2020	1.2250	1.6203	1.5670
carspcap	4.4517	4.3808	3.9764	3.8321	4.2280	4.0749
radiopcap	0.3214	0.3172	0.5860	0.6091	0.8314	0.7813
retailpcap	3.4621	3.4153	2.9609	2.8453	3.0229	3.0109
psinglem30_34	2.7225	2.7065	2.7108	2.7998	2.6958	2.6423

```
> oores <- ores - res
> apply(oores, 2, mad)

      MoranN      MoranR Prop_unstdN Prop_unstdR      Prop_stdN      Prop_stdR
0.03626358 0.02883196 0.04297479 0.04716456 0.03294942 0.02561857

> alpha_0.05 <- qnorm(0.05, lower.tail = FALSE)
> all((res >= alpha_0.05) == (ores >= alpha_0.05))
```

```
[1] TRUE
```

These assumptions affect the shape of the distribution of the measure in its tails; one possibility is to use a Saddlepoint approximation to find an equivalent to the analytical or bootstrap-based standard deviate for inference (Tiefelsdorf, 2002). The Saddlepoint approximation requires the eigenvalues of the weights matrix and iterative root-finding for global Moran's I , while for local Moran's I_i , analytical forms are known. Even with this computational burden, the Saddlepoint approximation for global Moran's I runs quite quickly. First we need to fit null linear models (only including an intercept) to the variables, then apply `lm.morantest.sad` to the fitted model objects:

```
> lm_objs <- lapply(vars, function(x) lm(formula(paste(x, "~1")), data = eire_ge1))
> system.time({
+   MoranSad <- lapply(lm_objs, function(x) lm.morantest.sad(x, listw = nb_B))
+   Prop_unstdSad <- lapply(lm_objs, function(x) lm.morantest.sad(x,
+     listw = lw_unstand))
+   Prop_stdSad <- lapply(lm_objs, function(x) lm.morantest.sad(x, listw = lw_std))
+ })

      user  system elapsed
0.386    0.001    0.397

> res <- sapply(c("MoranSad", "Prop_unstdSad", "Prop_stdSad"), function(x) sapply(get(x),
+   "[", "statistic"))
> rownames(res) <- vars
```

Although the analytical standard deviates (under Normality) are larger than those reached using the Saddlepoint approximation when measured by median absolute deviation, the differences do not lead to different inferences at this chosen cut-off. This reflects the fact that the shape of the distribution is very sensitive to small n , but for moderate n and global Moran's I , the effects are seen only further out in the tails. The consequences for local Moran's I_i are much stronger, because the clique of neighbours of each observation is typically very small. It is perhaps of interest that the differences are much smaller for the case of general weights than for unstandardised binary weights.

```
> print(formatC(res, format = "f", digits = 4), quote = FALSE)
```

	MoranSad	Prop_unstdSad	Prop_stdSad
pagval2_10	3.2903	3.1711	3.8283
ln_pagval2_10	3.5346	3.3982	4.0441
pagval10_50	1.0883	1.3219	1.4791
pagval50p	4.4402	4.4258	4.9377
cowspacre	4.4366	3.9164	4.7406
ln_cowspacre	4.4758	3.9760	4.6493
ocattlepacre	0.6030	2.0748	1.8642
pigspacre	2.2611	2.7154	3.0775
ln_pigspacre	2.1158	2.4271	2.7417
sheeppacre	1.0250	1.7040	1.4476
ln_sheeppacre	1.2669	2.2921	1.9731
townvillp	2.0949	1.2079	1.5872
carspcap	3.8500	3.6840	4.1044
radiopcap	0.3750	0.6094	0.7906
retailpcap	3.0663	2.8049	2.9245
psinglem30_34	2.4649	2.6449	2.6089

```
> oores <- res - ores[, c(1, 3, 5)]
> apply(oores, 2, mad)
```

MoranSad	Prop_unstdSad	Prop_stdSad
0.37142684	0.10779060	0.05650183

```
> all((res >= alpha_0.05) == (ores[, c(1, 3, 5)] >= alpha_0.05))
```

```
[1] TRUE
```

In addition we could choose to use the exact distribution of Moran's I , as described by Tiefelsdorf (2000); its implementation is covered in Bivand, Müller, and Reder (2009). The global case also needs the eigenvalues of the weights matrix, and the solution of a numerical integration function, but for these cases, the timings are quite acceptable.

```
> system.time({
+   MoranEx <- lapply(lm_objs, function(x) lm.morantest.exact(x, listw = nb_B))
+   Prop_unstdEx <- lapply(lm_objs, function(x) lm.morantest.exact(x,
+     listw = lw_unstand))
+   Prop_stdEx <- lapply(lm_objs, function(x) lm.morantest.exact(x,
+     listw = lw_std))
+ })

      user system elapsed
0.495   0.004   0.515

> res <- sapply(c("MoranEx", "Prop_unstdEx", "Prop_stdEx"), function(x) sapply(get(x),
+   "[", "statistic"))
> rownames(res) <- vars
```

The output is comparable with that of the Saddlepoint approximation, and the inferences drawn here are the same for the chosen cut-off as for the analytical standard deviates calculated under Normality.

```
> print(formatC(res, format = "f", digits = 4), quote = FALSE)
```

	MoranEx	Prop_unstdEx	Prop_stdEx
pagval2_10	3.2895	3.1660	3.8261
ln_pagval2_10	3.5384	3.3979	4.0430
pagval10_50	1.0798	1.3131	1.4745
pagval50p	4.4568	4.4430	4.9446
cowspacre	4.4532	3.9268	4.7453
ln_cowspacre	4.4928	3.9875	4.6531
ocattlepacre	0.5967	2.0611	1.8596
pigspacre	2.2486	2.7033	3.0740
ln_pigspacre	2.1031	2.4131	2.7380
sheeppacre	1.0168	1.6924	1.4430
ln_sheeppacre	1.2575	2.2779	1.9685
townvillp	2.0822	1.1999	1.5826
carspcap	3.8593	3.6899	4.1037
radiopcap	0.3696	0.6054	0.7873
retailpcap	3.0616	2.7938	2.9210
psinglem30_34	2.4533	2.6321	2.6050

```
> oores <- res - ores[, c(1, 3, 5)]
> apply(oores, 2, mad)
```

	MoranEx	Prop_unstdEx	Prop_stdEx
	0.37187539	0.09751723	0.05419300

```
> all((res >= alpha_0.05) == (ores[, c(1, 3, 5)] >= alpha_0.05))
```

```
[1] TRUE
```

Li, Calder, and Cressie (2007) take up the challenge in Cliff and Ord (1969, p. 31), to try to give the statistic a bounded fixed range. Their APLE measure is intended to approximate the spatial dependence parameter of a simultaneous autoregressive model better than Moran's I , and re-scales the measure by a function of the eigenvalues of the spatial weights matrix. APLE requires the use of row standardised weights.

```
> vars_scaled <- lapply(vars, function(x) scale(eire_ge1[[x]], scale = FALSE))
> nb_W <- nb2listw(lw_unstand$neighbours, style = "W")
> pre <- spdep::preAple(0, listw = nb_W)
```

```

> MoranAPPLE <- sapply(vars_scaled, function(x) spdep::inAple(x, pre))
> pre <- spdep::preAple(0, listw = lw_std, override_similarity_check = TRUE)
> Prop_stdAPPLE <- sapply(vars_scaled, function(x) spdep::inAple(x, pre))
> res <- cbind(MoranAPPLE, Prop_stdAPPLE)
> colnames(res) <- c("APPLE W", "APPLE Gstd")
> rownames(res) <- vars

```

In order to save time, we use the two internal functions `spdep::preAple` and `spdep::inAple`, since for each definition of spatial weights, the same eigenvalue calculations need to be made. The notation using the `::` operator says that the function with named after the operator is to be found in the namespace of the package named before the operator. The APPLE values repeat the pattern that we have already seen — for some variables, the measured autocorrelation is very similar irrespective of spatial weights definition, while for others, the change in the definition from binary to general does make a difference.

```

> print(formatC(res, format = "f", digits = 4), quote = FALSE)

```

	APPLE W	APPLE Gstd
pagval2_10	0.7702	0.6628
ln_pagval2_10	0.7615	0.6655
pagval10_50	0.3954	0.3446
pagval50p	0.7519	0.7313
cowspacre	0.8329	0.7408
ln_cowspacre	0.8148	0.7487
ocattlepacre	0.1468	0.4092
pigspacre	0.6227	0.6205
ln_pigspacre	0.5582	0.5887
sheeppacre	0.1594	0.2841
ln_sheeppacre	0.2046	0.4550
townvillp	0.3442	0.2644
carspcap	0.7140	0.6166
radiopcap	0.0083	0.1376
retailpcap	0.6376	0.5307
psinglem30_34	0.4094	0.4889

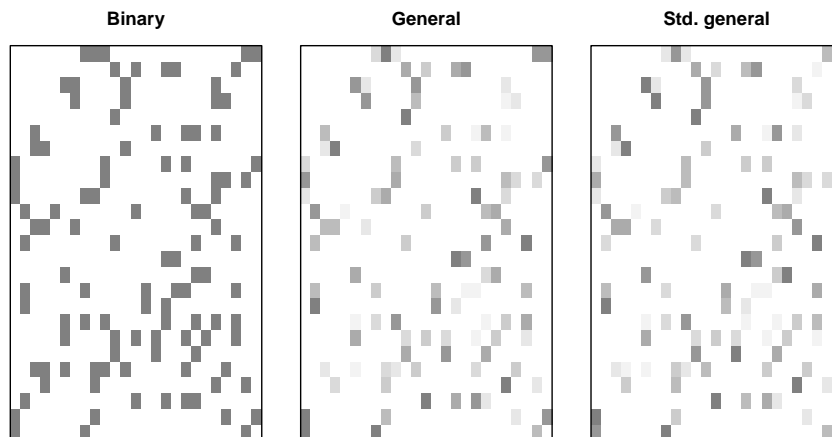


Figure 2: Three contrasted spatial weights definitions.

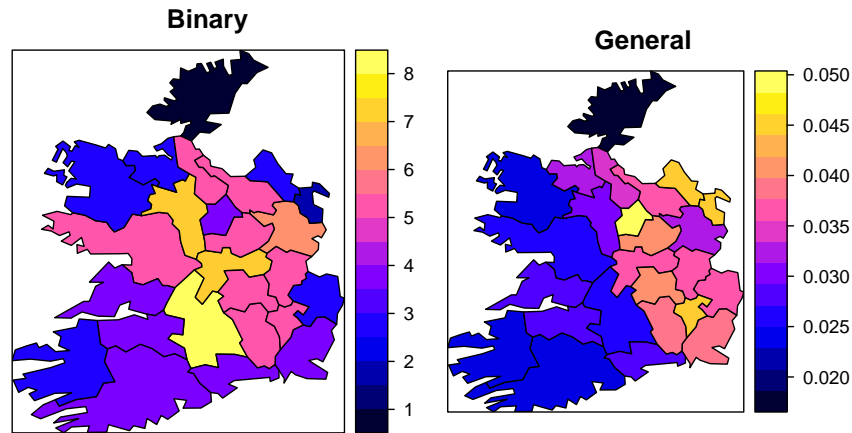


Figure 3: Sums of weights by county for two contrasted spatial weights definitions — for row standardisation, all counties sum to unity.

5 Odds and ends . . .

The differences found in the case of a few variables in inference using the original binary weights, and the general weights proposed by Cliff and Ord (1969) are necessarily related to the the weights themselves. Figures 2 and 3 show the values of the weights in sparse matrix form, and the sums of weights by county where these sums are not identical by design. In the case of binary weights, the matrix entries are equal, but the sums up-weight counties with many neighbours.

General weights up-weight counties that are close to each other, have more neighbours, and share larger boundary proportions (an asymmetric relationship). There is a further impact of using boundary proportions, in that the boundary between the county and the exterior is subtracted, thus boosting the weights between edge counties and their neighbours, even if there are few of them. Standardised general weights up-weight further up-weight counties with few neighbours, chiefly those on the edges of the study area.

With a small data set, here with $n = 25$, it is very possible that edge and other configuration effects are relatively strong, and may impact inference in different ways. The issue of egde effects has not really been satisfactorily resolved, and should be kept in mind in analyses of data sets of this size and shape.

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